

# **OPM-LMH SFM1**

#### Serum-free Cell Culture Medium

— For Vaccine Production

**OPM-LMH SFM1** is a serum-free cell culture medium designed for suspension culture of lmh cells and contains L-Glutamine. This medium supports the production of fowl adenovirus vaccine.

#### **Application**

OPM-LMH SFM1 is intended for large scale manufacturing of therapeutic biomolecules, as well as for research purposes, but not for human or any therapeutic use.

#### Storage & Transportation

#### **Shelf Life**

OPM-LMH SFM1 Dry Powder: 24 months OPM-LMH SFM1 Medium Liquid: 12 months

#### Reconstitution Method for Dry Powder

- 1. Measure out 90% of final required volume of purified water intended for cell culture use, e.g. WFI. Recommended water temperature is  $25\sim35^{\circ}$ C (minimum final volume  $\geq 1$ L).
- 2. Slowly add dry powder medium at 20.21 g/L and stir for 20 minutes.
- 3. Add 2.22 g/L NaHCO3 to the solution and continue to stir for 20 minutes.
- 4. Adjust pH to 8.5 with 5N NaOH and continue to stir until completely dissolved.
- 5. Adjust pH to 7.0 with 5N HCl.
- 6. Add cell culture grade purified water to 100% final volume.
- 7. Continue to stir for 10 minutes. Sterile filter using a membrane filter with a pore size of 0.22 micron.

#### **Quality Specifications**

Specifications	OPM-LMH SFM1 DPM	
Appearance	Khaki powder	
рН	7.0~7.5	
Osmolality (mOsm/kg)	250~300	
Solubility	Good by following the reconstitution instructions	
Endotoxin (EU/mL)	<2.0	

#### **Cell Culture Conditions**

37°C, 80% humidity, 5~8%CO₂

Shaker speed 110-150 rpm (amplitude: 50mm).



#### **Cell Recovery**

- 1. Rapidly thaw (<2 min) a vial of frozen cells in a 37 °C water bath.
- Transfer the entire contents aseptically into a 125 mL shake flask containing 30 mL prewarmed OPM-LMH SFM1 cell culture medium.
- 3. Incubate at 37 °C in a humidified atmosphere of  $5\%\sim8\%$  CO<sub>2</sub> in air on a shaker (rotating at  $110\sim130$  rpm (amplitude: 50mm).
- 4. Passage the cells for at least twice until fully recovery. Proceed according normal procedure after the Population Doubling Time stays stable.

## Cell Culture Passaging

- 1. Prewarm OPM-LMH SFM1 cell culture medium at 37  $^{\circ}$ C for 20~30min.
- 2. Proceed if VCD ≥1×10<sup>6</sup>/mL & viability ≥90%. Cultures should be passaged during the mid-log phase.
- Determine the correct volume of cell culture to inoculate a new flask at a starting cell density of 0.5×10<sup>6</sup> cells/mL in prewarmed OPM-LMH SFM1 cell culture medium
- 4. Incubate flasks in a humidified 37 °C incubator with 5%~8% CO₂ on an orbital shaker at 110~150rpm (amplitude: 50mm).
- 5. Passage cells by repeating the above steps every 2~3 days.

#### **Medium Adaptation**

#### **Direct Medium Adaptation**

- Cell lines may be adapted directly from serum-free media into OPM-LMH SFM1 cell culture medium.
  The seeding cell density can be referred to the passaging instructions or should be determined
  individually.
- 2. Cells should be passaged for a few times.
- 3. Adaptation is completed when the cultures attain stable VCD of  $2\times10^6$ /mL and viability  $\geq 90\%$  within  $3\sim4$  days over at least  $2\sim3$ passages.

#### Sequential Medium Adaptation

- For certain cell lines cultured in serum-free media, or in presence of 5~10% serum, sequential
  adaptation method is recommended.
- 2. Monitor the cell growth until the cell density has reached  $\geq 2 \times 10^6$  cells/mL.
- 3. Dilute the cells with a ratio of 25:75 (OPM-LMH SFM1 vs current medium), and then further dilute the culture until the cells grow well under this condition. Increase the proportion of OPM-LMH SFM1 in each subsequent operation, as is shown in the table.
- 4. Adaptation is completed when the cultures in 100% OPM-LMH SFM1 Cell culture medium attain stable VCD of 2×10<sup>6</sup>/mL and viability ≥ 85% within 3~4 days over at least 2~3 passages.

OPM-LMH SFM1: current medium (%)	Seeding density (×10 <sup>5</sup> cells/mL)	Evaluation of cell growth	Acceptance criteria for next step
25 : 75	3 ~ 4	VCD & Viability	VCD≥2×10 <sup>6</sup> /mL, Viability≥90% over 2 passages
50 : 50	3 ~ 4	VCD & Viability	VCD≥2×10 <sup>6</sup> /mL, Viability≥90% over 2 passages
75 : 25	3~4	VCD & Viability	VCD≥2×10 <sup>6</sup> /mL, Viability≥90% over 2 passages
90 : 10	3 ~ 4	VCD & Viability	VCD≥2×10 <sup>6</sup> /mL, Viability≥90% over 2 passages
100 : 0	3 ~ 4	VCD & Viability	VCD≥2×10°/mL, Viability≥90% over 2 passages



## Cryopreservation

- 1. Harvest the desired quantity of cells in mid-log phase of growth with viability over 90%.
- 2. Determine VCD to ensure that the final cell density is  $> 1 \times 10^7 / \text{ml}$ .
- 3. Prepare the freezing medium consisting of 90% OPM-LMH SFM1 Cell culture medium and 10% dimethyl sulfoxide (DMSO). Let the freezing medium cool down to  $4^{\circ}$ C.
- 4. Harvest cells by centrifugation at 400xg for 5 minutes. Remove the supernatant and resuspend the cell pellet with the cold freezing medium at  $> 1 \times 10^7/\text{ml}$ .
- 5. Transfer the suspension to sterile cryo-vials.
- 6. Place the vials in a cryo-box or a controlled rate freezing apparatus following standard procedures (1°C decrease per minute).
- 7. For long-term storage, transfer the vials to liquid nitrogen.



## **Order Information**

## Cell Culture Media

Name	Cat No.	Туре	Volume
OPM-LMH SFM1 DPM	C679018	Dry powder	50L/100L/500L
OPM-LMH SFM1 Medium	C221012	Liquid	1000mL



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